

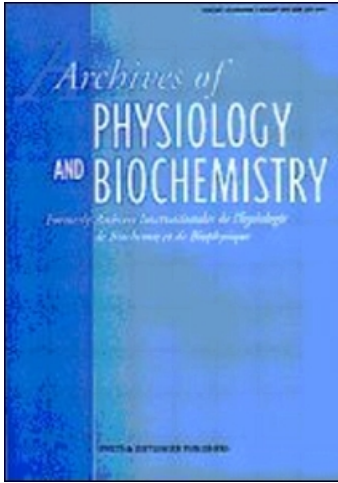
This article was downloaded by: [MALMAD]

On: 17 November 2008

Access details: *Access Details: [subscription number 902013951]*

Publisher *Informa Healthcare*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Archives Of Physiology And Biochemistry

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713817673>

Biology of peripheral blood cells in obstructive sleep apnea - the tip of the iceberg

Lena Lavie ^a; Larissa Dyugovskaya ^a; Andrey Polyakov ^a

^a Lloyd Rigler Sleep Apnea Research Laboratory, Unit of Anatomy and Cell Biology, The Ruth and Bruce Rappaport Faculty of Medicine, Technion-Israel Institute of Technology, Haifa, Israel

Online Publication Date: 01 October 2008

To cite this Article Lavie, Lena, Dyugovskaya, Larissa and Polyakov, Andrey(2008)'Biology of peripheral blood cells in obstructive sleep apnea - the tip of the iceberg',Archives Of Physiology And Biochemistry,114:4,244 — 254

To link to this Article: DOI: 10.1080/13813450802306701

URL: <http://dx.doi.org/10.1080/13813450802306701>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

REVIEW ARTICLE

Biology of peripheral blood cells in obstructive sleep apnea – the tip of the iceberg

LENA LAVIE, LARISSA DYUGOVSKAYA, & ANDREY POLYAKOV

Lloyd Rigler Sleep Apnea Research Laboratory, Unit of Anatomy and Cell Biology, The Ruth and Bruce Rappaport Faculty of Medicine, Technion-Israel Institute of Technology, Haifa, Israel

Abstract

Obstructive sleep apnea (OSA), a highly prevalent breathing disorder in sleep, characterized by intermittent and recurrent pauses in respiration, has emerged as an independent risk factor for cardiovascular morbidity and mortality. Accumulated evidence implicates Leukocyte-endothelial cell activation and adhesion as critical components that induce inflammation and injury to the vasculature resulting in the development of cardiovascular complications. Similar cellular interactions were described in conditions of ischemia/reperfusion, and various components of the metabolic syndrome as hypercholesterolemia and hypertension. The hallmark of sleep apnea – the multiple cycles of hypoxia/reoxygenation – promote oxidative stress and inflammation. These facilitate increased interactions of blood cells with endothelial cells, resulting in endothelial cell injury and dysfunction. Such events can promote atherosclerosis and the development of cardiovascular morbidities in OSA. However, inter-individual differences in response to intermittent hypoxia and activation of anti-inflammatory cytokine profiles in T lymphocytes can serve as protective mechanisms.

Key words: *Obstructive sleep apnea, inflammation, leukocytes, platelets, erythrocytes, endothelial cells.*

Introduction

Sleep medicine has witnessed an unprecedented growth in the last two decades due to the growing awareness of obstructive sleep apnea (OSA) and its profound impact on patients' quality of life and health (Tachibana *et al.*, 2005). OSA is a prevalent syndrome characterized by recurrent pauses in respiration, which result in cyclic decreases in blood oxygen saturation and sleep fragmentation. At least 4% of men and 2% of women are affected by OSA and display at least five disordered breathing events in each hour of sleep in the form of apneas or hypopneas (Punjabi, 2008; Young *et al.*, 1993). Apneas are complete cessations of breathing for at least 10 sec duration, while hypopneas are partial cessations for the same duration [defined as apnea-hypopnea-index (AHI) denoting the number of breathing arrests per hour of sleep]. These intermittent breathing arrests during sleep which are the hallmark of OSA syndrome are considered analogous to hypoxia/reperfusion injury. However, several characteristic symptoms such as excessive daytime

sleepiness, chronic fatigue or neurocognitive decline are an integral part as well (Young *et al.*, 1993). Specifically, the syndrome is associated with the male gender, middle age, central obesity, smoking, sedentary life-style, and a post-menopausal status in women (Young *et al.*, 2002b). However, the most dramatic impact is on the cardiovascular system (Lavie *et al.*, 2000; Smith *et al.*, 2002), particularly in relatively young populations of 50 years or younger (Lavie *et al.*, 2005). In recent years OSA was also shown to be associated with many risk factors of the metabolic syndrome such as hyperlipidemia, insulin resistance, hypertension and obesity. These may constitute additional risk factors for cardiovascular morbidity in OSA by acting synergistically with the apnoeic events to increase cardiovascular risk (Newman *et al.*, 2001; McArdle *et al.*, 2007; Punjabi *et al.*, 2002; Reichmuth *et al.*, 2005; Lam & Ip, 2007; Meslier *et al.*, 2003; Peppard *et al.*, 2000; Young *et al.*, 2002a; Basta & Vgontzas 2007).

The association between OSA and cardiovascular morbidity has been attributed to several mechanisms as increased activation of the sympathetic nervous

system, swings in intrathoracic pressure, altered blood coagulability, and, more recently to oxidative stress and inflammation (Carleson *et al.*, 1993; Shamsuzzaman *et al.*, 2003; Lavie, 2003). Of these, oxidative stress and inflammatory responses, which are fundamental mechanisms underlying atherosclerosis, are by far the most attractive proposed. Oxidative stress and subsequently inflammation and the acceleration of atherogenic processes are uniquely triggered by the apnea-related intermittent hypoxia (Lavie, 2003; Suzuki *et al.*, 2006). Moreover, these mechanisms are also integral parts of obesity, hypertension, and type 2 diabetes (Hotamisligil, 2006; Rudich *et al.*, 1998; Lassègue & Griendling, 2004; Stokes & Granger, 2005; Meigs *et al.*, 2007; Furukawa *et al.*, 2004). These involve activation of complex pathways and cellular interactions which culminate in endothelial dysfunction as a prelude to atherosclerosis.

The current review outlines the major epidemiological studies pointing to increased cardiovascular morbidity and associated risk factors which cluster with sleep apnea. The second part summarizes the current literature on phenotypic and functional changes and the inflammatory responses of various blood cells. The interactions of leukocytes/endothelial cells and the possible cellular mechanisms which are initiated by the breathing arrests during sleep are described, as well as their possible involvement in the development of cardiovascular morbidity via endothelial cell injury and dysfunction.

OSA and cardiovascular morbidity

The evidence supporting the association of OSA with cardiovascular morbidity is strong. At the time of diagnosis, at least 50% of the sleep apnea patients are hypertensive and 10–15% have cardiovascular disease including myocardial infarction, and stroke (Lavie *et al.*, 2000; Smith *et al.*, 2002). On the other hand, breathing disorders in sleep are prevalent among non-selected patients with cardiovascular diseases (Peker *et al.*, 1999; Moe *et al.*, 1996). Cross-sectional prospective and population based studies support the association between cardiovascular morbidity and breathing disorders in sleep demonstrating that this association is independent of major possible confounding cardiovascular risk factors (Peppard *et al.*, 2000; Nieto *et al.*, 2000; Bixler *et al.*, 2000; Young *et al.*, 1997). Several studies also demonstrated that OSA is an independent risk factor for cardiovascular mortality (Lavie, 2008; Lavie *et al.*, 1995; Yaggi *et al.*, 2005), and that effective treatment of the syndrome significantly decreased mortality (Marti *et al.*, 2002; Marin *et al.*, 2005).

The evidence supporting a causal relationship between sleep apnea and hypertension is particularly strong. It is based on large epidemiological studies (Lavie *et al.*, 2000; Nieto *et al.*, 2000; Bixler *et al.*, 2000; Young *et al.*, 1997) and prospective studies

(Peppard *et al.*, 2000). Furthermore, treatment with nasal continuous positive airway pressure (nCPAP) that ameliorated the apneas also lowered blood pressure (see a recent review in Bazzano *et al.*, 2007). These data on OSA as an independent risk factor for cardiovascular morbidity and the evidence on the mortality risk in these patients are summarized in recent reviews (McNicholas *et al.*, 2007; Lavie, 2007).

The obesity epidemic which emerged in the last two decades as one of the major cardiovascular risk factors is also associated with OSA. In many epidemiological, cross-sectional, clinic-based and population-based studies visceral obesity, in particular, was positively correlated with cardiovascular disease in these patients (Young *et al.*, 2002a; Basta & Vgontzas 2007). Notably, between 60% and 90% of OSA patients are obese (Anstead & Phillips, 1999). Similarly to OSA, obesity is associated with the male gender, post menopausal status in women, cardiovascular morbidity, hypertension, stroke, insulin resistance and type 2 diabetes (Kopelman, 2000; Wisse *et al.*, 2007). Although the nature of this association in OSA is not clear, the severity of OSA was shown to be aggravated by gaining weight, and improvement was noted under massive weight reduction by a controlled diet (Smith *et al.*, 1985) or by surgical means (Charuzi *et al.*, 1985).

Similarly, the prevalence of dyslipidemia, insulin resistance, glucose tolerance and type 2 diabetes, which constitute major risk factors for cardiovascular morbidity, were also found to be high among patients with OSA. These data on the associations of OSA with these cardiovascular risk factors are summarized in several reviews and are beyond of the scope of the current paper (Young *et al.*, 2002a; McNicholas *et al.*, 2007). All in all, there is sufficient data to suggest that cardiovascular morbidities and the associated metabolic syndrome risk factors are prevalent among patients with OSA.

Endothelial dysfunction and atherosclerosis in OSA

A large body of evidence implicates endothelial dysfunction in various conditions including the metabolic syndrome and sleep apnea. While in the normal state the endothelium regulates the vascular tone and interactions between the vessel wall and circulating substances and blood cells, endothelial dysfunction promotes an activated state. As a result, the anti-coagulant and anti-inflammatory endothelium acquires a pro-thrombotic and pro-inflammatory phenotype which initiates atherosclerosis (Libby, 2002; Davignon & Ganz, 2004). In endothelial dysfunction, the vasodilatation of the blood vessels is compromised due to decreased nitric oxide (NO) bioavailability. The impact of NO relies on its vasodilatory and protective properties which limit leukocyte recruitment and leukocyte expression of

adhesion molecules, and prevent the proliferation of vascular smooth muscle cells and aggregation/adhesion of platelets. Thus, it protects from the development of atherosclerosis. Exposure to oxidative stress, to inflammatory mediators or hypercholesterolemia, all of which promote endothelial cell activation and endothelial dysfunction, impair NO bioavailability (Libby, 2002; Davignon & Ganz, 2004; Lavie, 2004). In patients with OSA, several studies have demonstrated diminished NO bioavailability that was restored after nCPAP treatment (Schulz *et al.*, 2000b; Ip *et al.*, 2000; Lavie *et al.*, 2003). Moreover, several measures that represent early signs of atherosclerosis were shown to be elevated in OSA including increased intima-media thickness that was also severity dependent (Drager *et al.*, 2007; Minoguchi *et al.*, 2005; Drager *et al.*, 2005), arterial plaque formation (Kaynak *et al.*, 2003), calcified artery atheromas (Freidlander *et al.*, 1998), and higher pulse wave velocity (Nagahama *et al.*, 2004; Drager *et al.*, 2005). Thus, cardiovascular morbidity in sleep apnea is foreseeable. Moreover, the impact of OSA on endothelial dysfunction is further demonstrated by showing that such early signs of atherosclerosis can be improved after 3 or 4 months of treatment with nCPAP or dental device (Drager *et al.*, 2007; Itzhaki *et al.*, 2007).

Cellular and molecular mechanisms in OSA

Inflammatory pathway activation

Reactive oxygen species (ROS) and a state of oxidative stress are considered potent activators of inflammatory pathways. One of the more studied pathways of inflammatory cell activation is the upregulation of the transcription factor nuclear factor (NF) κ B, which is largely affected by oxidative stress. The activation of NF κ B induces increased expression of adhesion molecules and inflammatory cytokines in many cell types including leukocytes and endothelial cells (Lavie, 2008). Its up-regulation was demonstrated in polymorphonuclear leukocytes (PMNs) and monocytes of patients with OSA (Htoo *et al.*, 2006; Greenberg *et al.*, 2006; Yamauch *et al.*, 2006). Similar findings were also documented in a tissue culture model utilizing HeLa cells that were exposed to intermittent hypoxia *in vitro* (Ryan *et al.*, 2005). Moreover, intermittent hypoxia *in vitro* was shown to activate the NF κ B in an I kappa B kinase (IKK) dependent manner, at least in part, via activation of p38 mitogen activated protein kinase (MAPK) (Ryan *et al.*, 2007). Thus, the ROS molecules produced in response to intermittent hypoxia initiate a cascade of inflammatory pathways resulting in over-expression of adhesion molecules and pro-inflammatory cytokines. These adhesion molecules facilitate the recruitment and accumulation of leukocytes and platelets on the endothelial cells lining the vasculature and promote

PMNs/monocyte/lymphocyte/platelets/endothelial cells interactions. Such cellular interactions between blood cells and endothelial cells may result in injury to the endothelium (Lavie, 2008; Lavie, 2003).

Blood cell activation and expression of adhesion molecules

In the normal state circulating leukocytes and endothelial cells express basal levels of adhesion molecules. However, upon encounter with a variety of stimuli or insults including inflammation, infections, hypercholesterolemia, cytokines, hypoxia/re-oxygenation and sleep apnea, their expression is up-regulated. The expression of adhesion molecules is a highly regulated and sequential process and occurs in both endothelial cells and leukocytes. Up-regulated expression of adhesion molecules augments the interactions between these cell types and promotes the adherence of leukocytes to the vascular endothelium. Initially increased expression of selectins (L-selectins in leukocytes, E-selectins in endothelial cells and P-selectins in platelets and endothelial cells) facilitates weak binding of the leukocytes to endothelial cells. A firm binding is mediated by the integrins which also mediate transmigration into the interstitial layer through the endothelial cell layer (Libby, 2002; Panés & Granger, 1998). The interactions between endothelial cells and various leukocyte subpopulations of patients with OSA, including monocytes, polymorphonuclear leukocytes (PMNs), and numerous cytotoxic T cells expressing CD8, CD4, and $\gamma\delta$ molecules were rigorously investigated in our laboratory (Dyugovskaya *et al.*, 2002; Dyugovskaya *et al.*, 2005a; Dyugovskaya *et al.*, 2003; Dyugovskaya *et al.*, 2005b; Lavie *et al.*, 2005; Dyugovskaya *et al.*, 2008). A summary of the main phenotypic and functional changes of these cells in OSA is depicted in Table I.

Polymorphonuclear leukocytes (PMNs). The PMNs best known for their classical role as professional phagocytes are the most abundant of the leukocyte sub-populations, representing approximately 60% of all circulating leukocytes. They are short lived (up to 24 h in the blood stream) terminally differentiated cells that continuously undergo cell death by apoptosis. The constitutive apoptotic program controls and limits their life span and by that protects surrounding cells and tissues from their injurious compounds, i.e. ROS molecules, bactericidal proteins, lytic enzymes and leukotriens, which participate in inflammatory responses against invading micro-organisms, foreign particles or cellular debris. Another key feature of PMNs biology is their ability to express growth factors, inflammatory cytokines and chemokines, cell surface receptors, and adhesion molecules. Moreover, as they are the first to be recruited to inflammatory sites it is conceivable that they actively contribute to the sequential recruitment

Table I. The distribution of the main leukocyte adhesion molecules, markers, adhesion indices and reactive oxygen species (\pm SD) in fresh whole blood cells.

	Control	OSA	Cited by
<i>PMNs</i>			
CD15 (MFI)	1048 \pm 370	1484 \pm 470*	Dyugovskaya <i>et al.</i> (2008)
CD62L (MFI)	48.3 \pm 14.7	112.3 \pm 22.0*	Unpublished data
CD11b (MFI)	121.0 \pm 64.0	139.0 \pm 62.0 (ns)	Unpublished data
CD11c (%)	29.3 \pm 26.8	27.0 \pm 22.4 (ns)	Dyugovskaya <i>et al.</i> (2008)
Low-CD16 (%)	2.8 \pm 0.9	1.0 \pm 0.6*	Dyugovskaya <i>et al.</i> (2008)
Adhesion index	3.72 \pm 0.74	3.85 \pm 0.45 (ns)	Dyugovskaya <i>et al.</i> (2002)
ROS (MFI)	1079 \pm 355	1482 \pm 358*	Dyugovskaya <i>et al.</i> (2002)
Superoxide (nmoles/5 \times 10 ⁶ cells)	3.7 \pm 0.5	9.3 \pm 1.5*	Schulz <i>et al.</i> (2000a)
<i>Monocytes</i>			
CD15 (%)	1.4 \pm 0.9	12.4 \pm 7.4*	Dyugovskaya <i>et al.</i> (2002); Lavie <i>et al.</i> , (2005)
CD11c (%)	25.5 \pm 12.5	54.8 \pm 19.6*	Dyugovskaya <i>et al.</i> (2002)
Adhesion index	5.20 \pm 1.80	9.80 \pm 0.49*	Dyugovskaya <i>et al.</i> (2002)
ROS (MFI)	716 \pm 198	962 \pm 252*	Dyugovskaya <i>et al.</i> (2002)
<i>Lymphocytes</i>			
$\gamma\delta$ T cells/CD62L (%)	49.6 \pm 11	65.0 \pm 11.7*	Dyugovskaya <i>et al.</i> (2003)
$\gamma\delta$ T cells/adhesion index	3.60 \pm 0.90	6.10 \pm 0.90*	Dyugovskaya <i>et al.</i> (2003)
CD8+/CD40L+ (%)	3.4 \pm 2.5	14.4 \pm 10.5*	Dyugovskaya <i>et al.</i> (2005b)
CD8+/CD62L+ (%)	47.1 \pm 14.8	44.0 \pm 14.0 (ns)	Dyugovskaya <i>et al.</i> (2005b)
CD8+/adhesion index	3.54 \pm 1.40	3.90 \pm 0.90 (ns)	Unpublished data
CD4+/CD28 ^{null} (%)	6.8 \pm 1.3	19.9 \pm 5.0*	Dyugovskaya <i>et al.</i> (2005b)
CD4+/CD62L+ (%)	75.3 \pm 13.6	79.1 \pm 7.9 (ns)	Dyugovskaya <i>et al.</i> (2005b)
CD4+/adhesion index	3.47 \pm 1.4	3.90 \pm 2.40 (ns)	Unpublished data
ROS (MFI) – all lymphocytes	209 \pm 54	299 \pm 79*	Unpublished data

*Statistically significant, with $p < 0.05$ at least; ns, statistically not significant; MFI, mean fluorescence intensity, denoting intensity of expression; %, representing the percentage of cells from the population which express a specific molecule; ROS, reactive oxygen species. Adhesion index was determined on human umbilical vein endothelial cells (HUVECs) in culture.

of different leukocyte populations to inflammatory sites similarly to conditions characterized by ischemia and reperfusion (McDonald, 2004). Interestingly, PMNs were shown to infiltrate eroded or ruptured plaques obtained from patients with acute coronary syndromes (Naruko *et al.*, 2002; Zidar *et al.*, 2005) and to participate in the pathogenesis of lethal myocardial reperfusion (Vinten-Johansen, 2004). Their involvement in amplifying cardiovascular morbidity was further established by depletion experiments which resulted in reduced myocardial infarct size and a protected myocardium (Jolly *et al.*, 1986; Kin *et al.*, 2006).

Investigating PMNs' life span, functions and adhesive properties in patients with OSA revealed increased production of ROS molecules and attenuation by nCPAP (Schulz *et al.*, 2000a; Dyugovskaya *et al.*, 2002). Increased expression of selectins (of the family of adhesion molecules which mediate capture and tethering) CD62 and the CD15 (which is a carbohydrate complex on selectins) of PMNs was noted in OSA as well (Table I). The up-regulated CD15 expression in OSA was also severity dependent as attested by the number of apnoeic events. However, no differences were noted between OSA and controls up to 15 apnea-hypopnea events per hour. Treatment with nCPAP effectively lowered the expression of CD15 (Dyugovskaya *et al.*, 2008). Interestingly, the expression of CD11b (Vishnevsky *et al.*, unpublished observations) or CD11c, the β -subunits of the integrins (and counter receptors

for ICAM-1 on endothelial cells responsible for firm adhesion) was unaffected (Table I). This is not surprising given that adhesion of OSA PMNs to endothelial cells in vitro was also unaffected (Dyugovskaya *et al.*, 2002). The fact that selectins but not integrins of OSA patients PMNs were up-regulated implies that the interactions involving binding and tethering with endothelial cells are increased but not the firm adhesion. In addition PMN apoptosis, a fundamental injury-limiting mechanism and a key event in the control of inflammation, was also suppressed in OSA PMNs. Such suppressed apoptosis may imply that the PMNs/endothelial cells interactions initiated by the selectins could be exacerbated and by that amplify the destructive potential of PMNs towards the endothelium. It should be noted that the apneic events (AHI) but not sleep fragmentation were identified as an independent predictor of the percentage of apoptotic PMNs. Moreover, similar data were obtained in PMNs from healthy individuals exposed to experimental intermittent hypoxia in vitro (Dyugovskaya *et al.*, 2008). Thus, together with increased expression of selectins, suppressed PMNs apoptosis may promote endothelial injury in OSA.

Monocytes. Like PMNs, monocytes are best known for their classical role as professional phagocytes. However, unlike PMNs, monocytes are long lived and their initiation, participation in progression and persistence of atherosclerosis, is well established

(Libby, 2002, 2007). In inflammatory conditions they express adhesion molecules and release large quantities of ROS and inflammatory cytokines. Indeed ROS production by OSA monocytes, was increased as compared to controls (Table I). Apart from ROS, monocytes of patients with OSA were shown to express increased CD15 and CD11c adhesion molecules. Moreover, their increased expression of CD15 was dependent on the severity of the syndrome (Lavie *et al.*, 2005). Also, treatment of monocytes from healthy individuals with hypoxia *in vitro*, resulted in up-regulated CD15 expression. Unlike in PMNs, the CD11c integrin of OSA monocytes was also elevated, while treatment with nCPAP attenuated the levels of both CD15 and CD11c. Accordingly, increased adhesion of OSA monocytes was noted towards endothelial cells of venous (HUVEC) and arterial origin (HCAEC). By utilizing antibodies that neutralize selectins (anti-CD62) and integrins (anti-CD54) adhesion of monocytes to endothelial cells was attenuated to values comparable to controls (Dyugovskaya *et al.*, 2002). The involvement of monocytes in atherogenesis in OSA was further implicated by the observation that lipid uptake was increased in human macrophages that were treated with experimental intermittent hypoxia *in vitro* (Lattimore *et al.*, 2005).

T lymphocytes. The participation of various lymphocyte subpopulations was primarily implicated in atherogenesis through cytokine secretion and antibody production. Lymphocytes were shown to be prevalent in atherosclerotic lesions and to modulate atherosclerotic responses (Vanderlaan & Reardon, 2005; Song *et al.*, 2001). Natural killer (NK) lymphocytes, CD8+, CD4+, and $\gamma\delta$ T cells were all implicated in atherosclerotic sequelae, which further add to the complexity of atherosclerosis. Numerous T lymphocyte subpopulations were investigated in patients with OSA. Basically, all T cells studied in OSA (CD8+, CD4+, and $\gamma\delta$ T cells) express an activated and a cytotoxic phenotype. Assessment of $\gamma\delta$ T cells phenotype, and function revealed that expression of CD62L selectins was increased as compared to controls (Table I). Also adhesion to endothelial cells and cytotoxicity towards endothelial cells were higher in OSA. The higher avidity and cytotoxicity of OSA $\gamma\delta$ T cells were mainly attributed of the pro-inflammatory cytokine tumor-necrosis-factor- α (TNF- α). This since utilizing antibodies which neutralize TNF- α , abolished the cytotoxicity against endothelial cells. Cytotoxic receptors as CD56 did not seem to participate in this process in $\gamma\delta$ T cells since elimination of the CD56 bearing cells did not alter the cytotoxicity against endothelial cells (Dyugovskaya *et al.*, 2003).

Unlike in $\gamma\delta$ T cells, adhesion of CD4+ and CD8+ T cells to endothelial cells was unaffected by OSA (Table I). However, cytotoxicity towards

endothelial cells was increased in both CD8+ and CD4+ T cells of OSA patients. The killing abilities of CD8+ T lymphocytes were found to be apnea-hyponea-index (AHI) severity dependent. Yet, each subpopulation employed different killing mechanisms to damage endothelial cells (Dyugovskaya *et al.*, 2005a; Dyugovskaya *et al.*, 2005b; Dyugovskaya *et al.*, 2003). While endothelial cell killing by $\gamma\delta$ T lymphocytes was primarily mediated by TNF- α , CD8+ and CD4+ T lymphocytes utilized various other mechanisms. For instance, CD8+ T lymphocytes expressed more than three-fold of the CD40 ligand (CD40L) which is an important T cell activation marker and implicated in inflammatory cell activation in atherogenesis (Marx *et al.*, 2003). Additionally, OSA CD8+ T cells expressed higher amounts of the CD56 natural killer receptors and higher perforin levels which account for their higher cytotoxicity. Depletion of CD8+ T cells co-expressing CD56+ receptors greatly attenuated the cytotoxicity towards endothelial cells (Dyugovskaya *et al.*, 2005a). The CD4+ T cells of OSA patients contained about three-fold higher amounts of the subpopulation CD4+/CD^{28null} (Table I). These CD4+/CD^{28null} T cells are known to induce killing of endothelial cells (Liuzzo *et al.*, 2000). Yet it should be noted that the strongest cytotoxicity against endothelial cells was expressed by OSA $\gamma\delta$ T cells, CD8+ cytotoxicity was somewhat lower and that of CD4+ T cells was the lowest.

Platelets. Platelets are best known for their functions in maintaining vascular homeostasis by clot formation and wound healing. Under physiological conditions platelets circulate in a quiescent state protected from activation by inhibitory mediators released from intact endothelial cells, including NO. However, in response to endothelial dysfunction or when encountering vascular damage or under oxidative stress, platelets rapidly undergo activation, followed by interactions with monocytes and PMNs and increased adhesion and aggregation in the vessel wall (Zarbock *et al.*, 2007). This implicates their involvement in atherosclerosis. Similarly to circulating leukocytes, also platelets have been shown to acquire an activated and a pro-thrombotic phenotype in response to hypoxia/reoxygenation (Gavins *et al.*, 2007). Platelets from patients with OSA expressed increased activation and aggregability *in vitro*. The percentage of platelets expressing P-selectin (CD62P) was higher (Bokinsky *et al.*, 1995; Geiser *et al.*, 2002), mainly in the severe group of patients (Eisensehr *et al.*, 1998) and was effectively lowered by treatment with nCPAP (Hui *et al.*, 2004). In addition increases in hematocrit, blood viscosity, and fibrinogen, in patients with OSA could further affect hypercoagulability (Hoffstein *et al.*, 1994; Nobili *et al.*, 2000). Also, in an ongoing study in our lab, we found that platelets of patients with OSA form higher amounts of platelets/monocytes aggregates as

compared to their age, sex, and body mass index (BMI) matched controls (Vishnevsky *et al.*, unpublished observations). Owing to the fact that platelets were shown to play a key role in ischemic cardiovascular diseases, their altered activation state and hyper-aggregability may contribute to increased cardiovascular morbidity in OSA as well.

Collectively, the higher expression of adhesion molecules on platelets and leukocytes, the higher avidity and the ability to strongly attach to endothelial cells in culture conditions, the stronger cytotoxicity of T cells against endothelial cells, the delayed apoptosis expressed by PMNs, the higher ROS generated by monocytes and PMNs, and the higher aggregability of platelets are all markers of activation of the various blood cells investigated and can serve as markers of the possible ongoing processes that may damage the endothelium and initiate atherogenesis in patients with OSA.

Erythrocytes. The red blood cells (RBCs) constitute the major cell type in the circulation. Their primary role is the transport of oxygen to all tissues and organs. Under normal blood flow their adherence to endothelial cells is non-significant and their deformability facilitates tissue perfusion. Under hypoxic/ischemic conditions RBCs are capable of inducing and participating in inflammatory responses (Madjdpour *et al.*, 2003). However, which mechanisms are involved or whether they contribute to inflammation is unclear. Nonetheless, ROS molecules and redox sensitive transcription factors were proposed as major determinants. RBCs' adhesiveness and aggregation were shown to be elevated in cardiovascular risk factors as hypertension (Kesler *et al.*, 2006), atherosclerosis (Rotstein *et al.*, 2002) and obesity (Samocho-Bonet *et al.*, 2004). More recently, OSA was found to be associated with increased RBC aggregation/adhesion, which was correlated with an increase in the inflammatory marker CRP (Peled *et al.*, 2008). Being the major component in the circulation – the effects of hypoxia or intermittent hypoxia on RBCs functions and adhesive properties should be considered. This is particularly important in view of the fact that RBC aggregation/adhesion is also affected by various cardiovascular risk factors.

Activated endothelial cells. As with leukocytes and platelets, endothelial cell activation results in up-regulated expression of adhesion molecules and pro-inflammatory cytokines and promotes cellular interactions. In their non-activated state endothelial cells resist adhesion with leukocytes, platelets and RBCs. However, activation or injury by various factors as hypercholesterolemia, obesity, hypertension and hypoxia/re-oxygenation triggers the expression of adhesion molecules, which mediate these interactions (Gavins *et al.*, 2007; Packard & Libby, 2008). Until recently most findings on endothelial cell activation in OSA relied on indirect evidence. For instance, the presence of soluble variants of

adhesion molecules in the circulation of patients with OSA which originated from endothelial cells is indicative of their involvement. These variant adhesion molecules are shed when endothelial cells undergo activation. Several well established such molecules were identified including E-selectin and P-selectin which is stored in intracellular granules and released from activated platelets and endothelial cells (Zamarrón-Sanz *et al.*, 2006; Minoguchi *et al.*, 2007). Likewise, the intracellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1) were identified in the plasma of patients with OSA (Zamarrón-Sanz *et al.*, 2006; Ohga *et al.*, 1999; Ursavaş *et al.*, 2007; Chin *et al.*, 2000). These circulating adhesion molecules are regarded as markers of active atherosclerotic diseases, and as predictors of future cardiovascular disease (Lavie, 2005). Additionally, soluble P-selectin (sP-selectin) levels reported to be higher in OSA patients were also negatively correlated with PMN apoptosis. This indicates a possible involvement of P-selectin in the inhibition of PMN apoptosis as observed in OSA, which may exacerbate PMNs functions to injure the endothelium (Dyugovskaya *et al.*, 2008). Of note, activation of endothelial cells in vivo was recently confirmed in OSA by harvesting endothelial cells obtained from veins. In that study, the authors demonstrated that OSA directly affects the vascular endothelium by promoting oxidative stress and inflammation while decreasing NO bioavailability and repair capacity (Jelic *et al.*, 2008).

Pro-inflammatory cytokines

Pro-inflammatory cytokines, like adhesion molecules, are affected by the redox state of the cells in which they are synthesized and actively participate and modulate inflammatory responses. Various inflammatory cytokines are synthesized and released by inflammatory cells. These multi-purpose molecules regulate both the innate and adaptive immune system. Cytokines regulate macrophage activation via expression of scavenger receptors and secretion of metalloproteinases, modulate the proliferation of smooth muscle cells, the production of nitric oxide and apoptosis, and stimulate the activation of endothelial cells. All of which are steps in the progression of atherosclerosis. The main cytokines investigated in OSA include: Tumor necrosis factor- α (TNF- α) that affects the initiation and progression of cardiovascular pathology (von der Thüsen *et al.*, 2003; Ridker *et al.*, 2000), interleukin-6 (IL-6), interleukin-8 (IL-8) and the anti-inflammatory cytokine IL-10 which were all shown to be affected by OSA. Evidently, once the inflammatory response is initiated, these cytokines can in turn activate NF κ B and by that can further exaggerate inflammation. In patients with OSA, increases in pro-inflammatory cytokines levels were primarily found in the circulation (Vgontzas *et al.*, 2000; Constantinidis

et al., 2008). But elevated levels in monocytes (Golan-Shany *et al.*, unpublished observations) and in various cytotoxic T lymphocytes were also observed (Dyugovskaya *et al.*, 2005a; Dyugovskaya *et al.*, 2003). Specifically, in $\gamma\delta$ T lymphocytes the pro-inflammatory TNF- α was increased and the anti-inflammatory IL-10 was decreased whereas the opposite was noted in controls (Figure 1). Also, the expression of IL-8, a pro-inflammatory cytokine with strong chemoattractant and activating properties for PMNs was shown to increase in $\gamma\delta$ T cells of patients with OSA (Figure 1). This clearly attests to a pro-inflammatory state in these cells. In CD8+ T cells both TNF- α and IL-10 were increased. The percentage of CD8+ cells expressing TNF- α was increased by four-fold whereas an incremental increase of 1.3-fold was noted for IL-10 (Figure 2). By contrast, in

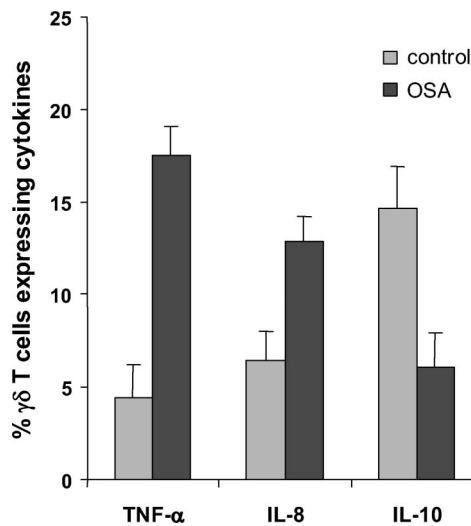


Figure 1. Percentage of $\gamma\delta$ T cells from sleep apnea patients and controls expressing the cytokines TNF- α , IL-8, and IL-10. Data adapted from Dyugovskaya *et al.* (2003).

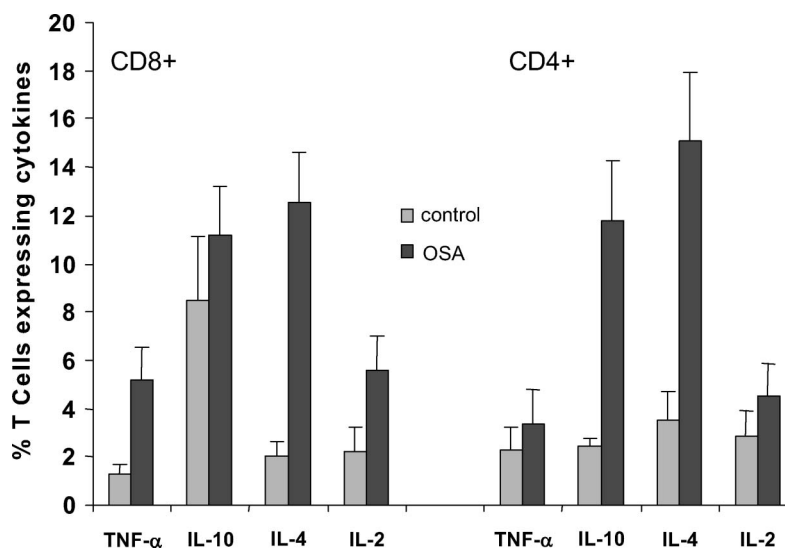


Figure 2. Percentage of CD8+ and CD4+ T cells from sleep apnea patients and controls expressing the cytokines TNF- α , IL-10, IL-4, and IL-2. Data adapted from Dyugovskaya *et al.* (2005b).

CD4+ T cells the percentage of cells expressing TNF- α was unaffected by OSA, but the expression of IL-10 was increased by 4.9-fold compared with control (Figure 2). Altered cytokine balance can result in activated T cells and can lead to their differentiation into effector cells with tissue damaging potential or with abilities to diminish inflammation. Depending on the cytokines produced upon such activation, T cells can be defined as type 1 – those that secrete IL-2 (and IFN- γ), and type 2 – those that secrete IL-4 (but also IL-5, IL-6 and IL-10). Based on the ratios of IL-2 and IL-4, CD4+ and CD8+ T cells of patients with OSA had increased percentage of IL-4 expressing cells than IL-2. This suggests a type 2-dominant cytokine expression, as demonstrated in Figure 2. This shift towards a type 2-dominance may represent a compensatory response of the CD8+ and CD4+ T cells to protect from other inflammatory cells or exacerbated pro-inflammatory responses. As both IL-10 and IL-4 have been shown to possess anti-inflammatory properties they can moderate inflammation (Libby, 2007). It should be stressed, however, that unlike the data presented here on cytokine expression of specific inflammatory/immune cells, the data on cytokines levels in OSA patients thus far were mostly acquired from serum or plasma. Such data represent the overall pool of the cytokines released from various inflammatory cells and adipocytes and therefore cannot delineate a specific inflammatory/anti-inflammatory response or an on-going process as described here for the cytokines investigated in the various cells. The cytokines released by adipocytes, in particular, can pose a problem. Since the fat tissue represents a major source for cytokines/adipokines, obese subjects express higher levels of inflammatory cytokines and low grade inflammation. Such data that were demonstrated in obese and overweight patients undergoing

surgical treatment (Constantinidis *et al.*, 2008) clearly attest to the need to separate the obesity component from the apneic events.

Conclusion and perspectives

Inflammatory cell activation is by now recognized as a fundamental mechanism in the pathophysiology of cardiovascular morbidity. Moreover, it is also involved in hypertension, hypercholesterolemia, glucose metabolism and obesity, which are all well established cardiovascular risk factors. In recent years the participation of inflammatory cell activation was recognized as an integral part of OSA pathophysiology as well. As specified before, intermittent hypoxia resulting from the apneas has profound effects on redox systems and initiates oxidative stress and inflammation via activation of NF κ B and downstream inflammatory/immune pathways. Consequently, up-regulated expression of adhesion molecules, inflammatory cytokines and ROS molecules, are evident in various leukocyte subpopulations and platelets of patients with OSA. These induce increased avidity and cytotoxicity towards endothelial cells. Increased erythrocyte aggregation was also noted in OSA, and should be considered as an important target of research. All in all, these cellular interactions in the vasculature promote endothelial dysfunction and early signs of atherosclerosis.

In view of the fact that many of these cellular events described are OSA severity dependent, it could be expected that all or most patients with OSA, in particularly those with severe OSA would suffer from various cardiovascular morbidities. This, however, is not the case. Even though cardiovascular morbidity and mortality in OSA is higher than in the general population, some OSA patients are free of cardiovascular morbidities suggesting the possible involvement of as yet unidentified protective mechanisms. Previously, we proposed that mechanisms as ischemic preconditioning inducing angiogenesis of newly formed heart collaterals may serve to protect some OSA patients (Lavie & Lavie, 2006). We also demonstrated large inter-individual differences in the angiogenic response to hypoxia that were correlated with arterial collateral formation in the heart. These may help to explain the inter-individual differences in cardiovascular morbidity (Schultz *et al.*, 1999). Here, we propose that CD8⁺ and even more so CD4⁺ T cells can participate in such protective mechanisms via shift to type 2 cytokine secretion and dominance, and by that moderate inflammation. Thus, there is a need to elucidate many basic unanswered questions for a better understanding of the sequence of interactions among all blood cells and endothelial cells, and of the cytokines produced by each cell type and the overall cytokine balance. Clarifying which additional adhesion molecules participate, what is the sequence of interactions among inflammatory cells and endothelial cells, do the leukocytes transmigrate

to atherosclerotic plaques after activation, or, which of the transcription factors or transduction mechanisms participate? These are only a few examples, and a better understanding of these mechanisms can help in the development of new treatment modalities to prevent cardiovascular morbidity in OSA.

Acknowledgements

This study was supported in part by a grant from the Binational US-Israel foundation (BSF) grant no. 1006695.

Declaration of interest: The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

References

- Anstead M, Phillips B. 1999. The spectrum of sleep-disordered breathing. *Respir Care Clin N Am* 5:363–77, viii.
- Basta M, Vgontzas AN. 2007. Metabolic abnormalities in obesity and sleep apnea are in a continuum. *Sleep Med* 8:5–7.
- Bazzano LA, Khan Z, Reynolds K, He J. 2007. Effect of nocturnal nasal continuous positive airway pressure on blood pressure in obstructive sleep apnea. *Hypertension* 50:417–23.
- Bixler EO, Vgontzas AN, Lin HM, Ten Have T, Leiby BE, Vela-Bueno A, Kales A. 2000. Association of hypertension and sleep-disordered breathing. *Arch Intern Med* 160:2289–95.
- Bokinsky G, Miller M, Ault K, Husband P, Mitchell J. 1995. Spontaneous platelet activation and aggregation during obstructive sleep apnea and its response to therapy with nasal continuous positive airway pressure. A preliminary investigation. *Chest* 108:625–30.
- Carlson JT, Hedner J, Elam M, Ejjnell H, Sellgren J, Wallin BG. 1993. Augmented resting sympathetic activity in awake patients with obstructive sleep apnea. *Chest* 103:1763–8.
- Charuzi I, Ovnat A, Peiser J, Saltz H, Weitzman S, Lavie P. 1985. The effect of surgical weight reduction on sleep quality in obesity-related sleep apnea syndrome. *Surgery* 97:535–8.
- Chin K, Nakamura T, Shimizu K, Mishima M, Nakamura T, Miyasaka M, Ohi M. 2000. Effects of nasal continuous positive airway pressure on soluble cell adhesion molecules in patients with obstructive sleep apnea syndrome. *Am J Med* 109:562–7.
- Constantinidis J, Erelidis S, Angouridakis N, Konstantinidis I, Vital V, Angouridaki C. 2008. Cytokine changes after surgical treatment of obstructive sleep apnea syndrome. *Eur Arch Otorhinolaryngol* [Epub ahead of print], PMID: 18317790.
- Davignon J, Ganz P. 2004. Role of endothelial dysfunction in atherosclerosis. *Circulation* 109(23 Suppl 1):III27–32.
- Drager LF, Bortolotto LA, Figueiredo AC, Krieger EM, Lorenzi-Filho G. 2007. Effects of continuous positive airway pressure on early signs of atherosclerosis in obstructive sleep apnea. *Am J Respir Crit Care Med* 176:706–12.
- Drager LF, Bortolotto LA, Lorenzi MC, Figueiredo AC, Krieger EM, Lorenzi-Filho G. 2005. Early signs of atherosclerosis in obstructive sleep apnea. *Am J Respir Crit Care Med* 172:613–8.
- Dyugovskaya L, Lavie P, Hirsh M, Lavie L. 2005a. Activated CD8⁺ T-lymphocytes in obstructive sleep apnea. *Eur Respir J* 25:820–8.
- Dyugovskaya L, Lavie P, Lavie L. 2002. Increased adhesion molecules expression and production of reactive oxygen species in leukocytes of sleep apnea patients. *Am J Respir Crit Care Med* 165:934–9.
- Dyugovskaya L, Lavie P, Lavie L. 2003. Phenotypic and functional characterization of blood gammadelta T cells in sleep apnea. *Am J Respir Crit Care Med* 168:242–9, 66.

- Dyugovskaya L, Lavie P, Lavie L. 2005b. Lymphocyte activation as a possible measure of atherosclerotic risk in patients with sleep apnea. *Ann NY Acad Sci* 1051:340–50.
- Dyugovskaya L, Polyakov A, Lavie P, Lavie L. 2008. Delayed neutrophil apoptosis in sleep apnea patients. *Am J Respir Crit Care Med* 177:544–54.
- Eisensehr I, Ehrenberg BL, Noachtar S, Korbett K, Byrne A, McAuley A, Palabrica T. 1998. Platelet activation, epinephrine, and blood pressure in obstructive sleep apnea syndrome. *Neurology* 51:188–95.
- Friedlander AH, Yueh R, Littner MR. 1998. The prevalence of calcified carotid artery atheromas in patients with obstructive sleep apnea syndrome. *J Oral Maxillofac Surg* 56:950–4.
- Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, Nakayama O, Makishima M, Matsuda M, Shimomura I. 2004. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* 114:1752–61.
- Gavins F, Yilmaz G, Granger DN. 2007. The evolving paradigm for blood cell-endothelial cell interactions in the cerebral microcirculation. *Microcirculation* 14:667–81.
- Geiser T, Buck F, Meyer BJ, Bassetti C, Haeberli A, Gugger M. 2002. In vivo platelet activation is increased during sleep in patients with obstructive sleep apnea syndrome. *Respiration* 69:229–34.
- Greenberg H, Ye X, Wilson D, Htoo AK, Hendersen T, Liu SF. 2006. Chronic intermittent hypoxia activates nuclear factor-kappaB in cardiovascular tissues in vivo. *Biochem Biophys Res Commun* 343:591–6.
- Hoffstein V, Herridge M, Mateika S, Redline S, Strohl KP. 1994. Hematocrit levels in sleep apnea. *Chest* 106:787–91.
- Hotamisligil GS. 2006. Inflammation and metabolic disorders. *Nature* 444:860–67.
- Htoo AK, Greenberg H, Tongia S, Chen G, Henderson T, Wilson D, Liu SF. 2006. Activation of nuclear factor kappaB in obstructive sleep apnea: a pathway leading to systemic inflammation. *Sleep Breath* 10:43–50.
- Hui DS, Ko FW, Fok JP, Chan MC, Li TS, Tomlinson B, Cheng G. 2004. The effects of nasal continuous positive airway pressure on platelet activation in obstructive sleep apnea syndrome. *Chest* 125:1768–75.
- Ip MS, Lam B, Chan LY, Zheng L, Tsang KW, Fung PC, Lam WK. 2000. Circulating nitric oxide is suppressed in obstructive sleep apnea and is reversed by nasal continuous positive airway pressure. *Am J Respir Crit Care Med* 162:2166–71.
- Itzhaki S, Dorchin H, Clark G, Lavie L, Lavie P, Pillar G. 2007. The effects of 1-year treatment with a Herbst mandibular advancement splint on obstructive sleep apnea, oxidative stress, and endothelial function. *Chest* 131:740–9.
- Jelic S, Padeletti M, Kawut SM, Higgins C, Canfield SM, Onat D, Colombo PC, Basner RC, Factor P, Lejemtel TH. 2008. Inflammation oxidative stress, and repair capacity of the vascular endothelium in obstructive sleep apnea. *Circulation* 117:2270–8.
- Jolly SR, Kane WJ, Hook BG, Abrams GD, Kunkel SL, Lucchesi BR. 1986. Reduction of myocardial infarct size by neutrophil depletion: effect of duration of occlusion. *Am Heart J* 112:682–90.
- Kaynak D, Goksan B, Kaynak H, Degirmenci N, Daglioglu S. 2003. Is there a link between the severity of sleep-disordered breathing and atherosclerotic disease of the carotid arteries? *Eur J Neurol* 10:487–93.
- Kesler A, Yatziv Y, Shapira I, Berliner S, Assayag EB. 2006. Increased red blood cell aggregation in patients with idiopathic intracranial hypertension. A hitherto unexplored pathophysiological pathway. *Thromb Haemostasis* 96:483–7.
- Kin H, Wang NP, Halkos ME, Kerendi F, Guyton RA, Zhao ZQ. 2006. Neutrophil depletion reduces myocardial apoptosis and attenuates NFkappaB activation/TNFalpha release after ischemia and reperfusion. *J Surg Res* 135:170–8.
- Kopelman PG. 2000. Obesity as a medical problem. *Nature* 404:635–43.
- Lam JC, Ip MS. 2007. An update on obstructive sleep apnea and the metabolic syndrome. *Curr Opin Pulm Med* 13:484–89.
- Lassègue B, Griendling KK. 2004. Reactive oxygen species in hypertension. An update. *Am J Hypertens* 17:852–60.
- Lattimore JD, Wilcox I, Nakhla S, Langenfeld M, Jessup W, Celermajer DS. 2005. Repetitive hypoxia increases lipid loading in human macrophages – a potentially atherogenic effect. *Atherosclerosis* 179:255–9.
- Lavie L, Dyugovskaya L, Lavie P. 2005a. Sleep apnea related intermittent hypoxia and atherogenesis: adhesion molecules and monocytes/endothelial cells interactions. *Atherosclerosis* 183:183–4.
- Lavie L, Hefetz A, Luboshitzky R, Lavie P. 2003. Plasma levels of nitric oxide and L-arginine in sleep apnea patients: effects of nCPAP treatment. *J Mol Neurosci* 21:57–63.
- Lavie L, Lavie P. 2006. Ischemic preconditioning as a possible explanation for the age decline relative mortality in sleep apnea. *Med Hypotheses* 66:1069–73.
- Lavie L. 2003. Obstructive sleep apnea syndrome – an oxidative stress disorder. *Sleep Med Rev* 7:35–51.
- Lavie L. 2004. Sleep apnea syndrome, endothelial dysfunction, and cardiovascular morbidity. *Sleep* 27:1053–5.
- Lavie L. 2005. Sleep-disordered breathing and cerebrovascular disease: a mechanistic approach. *Neurol Clin* 23:1059–75.
- Lavie L. 2008. Intermittent hypoxia: the culprit of oxidative stress, vascular inflammation and dyslipidemia in obstructive sleep apnea. *Expert Rev Resp Med* 2:75–84.
- Lavie P, Herer P, Hoffstein V. 2000. Obstructive sleep apnea syndrome as a risk factor for hypertension: population study. *BMJ* 320:479–82.
- Lavie P, Herer P, Peled R, Berger I, Yoffe N, Zomer J, Rubin AH. 1995. Mortality in sleep apnea patients: a multivariate analysis of risk factors. *Sleep* 18:149–57.
- Lavie P, Lavie L, Herer P. 2005b. All-cause mortality in males with sleep apnea syndrome: declining mortality rates with age. *Eur Respir J* 25:514–20.
- Lavie P. 2007. Mortality in sleep apnea syndrome: review of the evidence. *Eur Respir Rev* 16:203–10.
- Libby P. 2002. Inflammation in atherosclerosis. *Nature*. 420:868–74.
- Libby P. 2007. Inflammatory mechanisms: the molecular basis of inflammation and disease. *Nutr Rev* 65(12 Pt 2):S140–6.
- Liuzzo G, Goronzy JJ, Yang H, Kopecky SL, Holmes DR, Frye RL, Weyand CM. 2000. Monoclonal T-cell proliferation and plaque instability in acute coronary syndromes. *Circulation* 101:2883–8.
- Madjdipour C, Jewell UR, Kneller S, Ziegler U, Schwendener R, Booy C, Kläusli L, Pasch T, Schimmer RC, Beck-Schimmer B. 2003. Decreased alveolar oxygen induces lung inflammation. *Am J Physiol Lung Cell Mol Physiol* 284:L360–7.
- Marin JM, Carrizo SJ, Vicente E, Agusti AG. 2005. Long-term cardiovascular outcomes in men with obstructive sleep apnea-hypopnea with or without treatment with continuous positive airway pressure: an observational study. *Lancet* 365:1046–53.
- Marti S, Sampol G, Muñoz X, Torres F, Roca A, Lloberes P, Sagalés T, Quesada P, Morell F. 2002. Mortality in severe sleep apnea/hypopnea syndrome patients: impact of treatment. *Eur Respir J* 20:1511–18.
- Marx N, Imhof A, Froehlich J, Siam L, Ittner J, Wierse G, Schmidt A, Maerz W, Hombach V, Koenig W. 2003. Effect of rosiglitazone treatment on soluble CD40L in patients with type 2 diabetes and coronary artery disease. *Circulation* 107:1954–7.
- McArdle N, Hillman D, Beilin L, Watts G. 2007. Metabolic risk factors for vascular disease in obstructive sleep apnea: a matched controlled study. *Am J Respir Crit Care Med* 175:190–5.
- McDonald PP. 2004. Transcriptional regulation in neutrophils: teaching old cells new tricks. *Adv Immunol* 82:1–48.
- McNicholas WT, Bonsignore MR and Management Committee of EU COST ACTION B26. 2007. Sleep apnea as an independent risk factor for cardiovascular disease: current evidence, basic mechanisms and research priorities. *Eur Respir J* 29:156–78.

- Meigs JB, Larson MG, Fox CS, Keane JF Jr, Vasan RS, Benjamin EJ. 2007. Association of oxidative stress, insulin resistance, and diabetes risk phenotypes: the Framingham Offspring Study. *Diabetes Care* 30:2529–35.
- Meslier N, Gagnadoux F, Giraud P, Person C, Oukel H, Urban T, Racineux JL. 2003. Impaired glucose-insulin metabolism in males with obstructive sleep apnea syndrome. *Eur Respir J* 22:156–60.
- Minoguchi K, Yokoe T, Tazaki T, Minoguchi H, Oda N, Tanaka A, Yamamoto M, Ohta S, O'Donnell CP, Adachi M. 2007. Silent brain infarction and platelet activation in obstructive sleep apnea. *Am J Respir Crit Care Med* 175:612–7.
- Minoguchi K, Yokoe T, Tazaki T, Minoguchi H, Tanaka A, Oda N, Okada S, Ohta S, Naito H, Adachi M. 2005. Increased carotid intima-media thickness and serum inflammatory markers in obstructive sleep apnea. *Am J Respir Crit Care Med* 172:625–30.
- Moore T, Rabben T, Wiklund U, Franklin KA, Eriksson P. 1996. Sleep-disordered breathing in men with coronary artery disease. *Chest* 109:659–63.
- Nagahama H, Soejima M, Uenomachi H, Higashi Y, Yotsumoto K, Samukawa T, Arima T. 2004. Pulse wave velocity as an indicator of atherosclerosis in obstructive sleep apnea syndrome patients. *Intern Med* 43:184–8.
- Naruko T, Ueda M, Haze K, van der Wal AC, van der Loos CM, Itoh A, Komatsu R, Ikura Y, Ogami M, Shimada Y, *et al.* 2002. Neutrophil infiltration of culprit lesions in acute coronary syndromes. *Circulation* 106:2894–900.
- Newman AB, Nieto FJ, Guidry U, Lind BK, Redline S, Pickering TG, Quan SF. Sleep Heart Health Study Research Group. 2001. Sleep Heart Health Study Research Group. Relation of sleep-disordered breathing to cardiovascular disease risk factors: the Sleep Heart Health Study. *Am J Epidemiol* 154:50–59.
- Nieto FJ, Young TB, Lind BK, Shahar E, Samet JM, Redline S, D'Agostino RB, Newman AB, Lebowitz MD, Pickering TG. 2000. Association of sleep-disordered breathing, sleep apnea, and hypertension in a large community-based study. Sleep Heart Health Study. *JAMA* 283:1829–36.
- Nobili L, Schiavi G, Bozano E, De Carli F, Ferrillo F, Nobili F. 2000. Morning increase of whole blood viscosity in obstructive sleep apnea syndrome. *Clin Hemorheol Microcirc* 22:21–7.
- Ohga E, Nagase T, Tomita T, Teramoto S, Matsuse T, Katayama H, Ouchi Y. 1999. Increased levels of circulating ICAM-1, VCAM-1, and L-selectin in obstructive sleep apnea syndrome. *J Appl Physiol* 87:10–4.
- Packard RR, Libby P. 2008. Inflammation in atherosclerosis: from vascular biology to biomarker discovery and risk prediction. *Clin Chem* 54:24–38.
- Panés J, Granger DN. 1998. Leukocyte-endothelial cell interactions: molecular mechanisms and implications in gastrointestinal disease. *Gastroenterology* 114:1066–90.
- Peker Y, Kraiczki H, Hedner J, Löth S, Johansson A, Bende M. 1999. An independent association between obstructive sleep apnea and coronary artery disease. *Eur Respir J* 14:179–84.
- Peled N, Kassirer M, Kramer MR, Rogowski O, Shlomi D, Fox B, Berliner AS, Shitrit D. 2008. Increased erythrocyte adhesiveness and aggregation in obstructive sleep apnea syndrome. *Thromb Res* 121:631–6.
- Peppard PE, Young T, Palta M, Skatrud J. 2000. Prospective study of the association between sleep disordered breathing and hypertension. *N Engl J Med* 342:1738–84.
- Punjabi NM. 2008. The epidemiology of adult obstructive sleep apnea. *Proc Am Thorac Soc* 5:136–43.
- Punjabi NM, Sorkin JD, Katzell LJ, Goldberg AP, Schwartz AR, Smith PL. 2002. Sleep-disordered breathing and insulin resistance in middle-aged and overweight men. *Am J Respir Crit Care Med* 165:677–82.
- Reichmuth KJ, Austin D, Skatrud JB, Young T. 2005. Association of sleep apnea and type II diabetes: a population-based study. *Am J Respir Crit Care Med* 172:1590–95.
- Ridker PM, Rifai N, Pfeffer M, Sacks F, Lepage S, Braunwald E. 2000. Elevation of tumor necrosis factor-alpha and increased risk of recurrent coronary events after myocardial infarction. *Circulation* 101:2149–53.
- Rotstein R, Landau T, Twig A, Rubinstein A, Koffler M, Justo D, Constantiner D, Zeltser D, Shapira I, Mardi T, *et al.* 2002. The erythrocyte adhesiveness/aggregation test (EAAT). A new biomarker to reveal the presence of low grade subclinical smoldering inflammation in individuals with atherosclerotic risk factors. *Atherosclerosis* 165:343–51.
- Rudich A, Tirosh A, Potashnik R, Hemi R, Kanety H, Bashan N. 1998. Prolonged oxidative stress impairs insulin-induced GLUT4 translocation in 3T3-L1 adipocytes. *Diabetes* 47:1562–9.
- Ryan S, McNicholas WT, Taylor CT. 2007. A critical role for p38 map kinase in NF-kappaB signalign during intermittent hypoxia/reoxygenation. *Biochem Biophys Res Commun* 355:728–33.
- Ryan S, Taylor CT, McNicholas WT. 2005. Selective activation of inflammatory pathways by intermittent hypoxia in obstructive sleep apnea syndrome. *Circulation* 112:2660–7.
- Samocha-Bonet D, Ben-Ami R, Shapira I, Shenkerman G, Abu-Abied S, Stern N, Mardi T, Tulchinski T, Deutsch V, Yedgar S, *et al.* 2004. Flow-resistant red blood cell aggregation in morbid obesity. *Int J Obes Relat Metab Disord* 28:1528–34.
- Schultz A, Lavie L, Hochberg I, Beyar R, Stone T, Skorecki K, Lavie P, Roguin A, Levy AP. 1999. Interindividual heterogeneity in the hypoxic regulation of VEGF: significance for the development of the coronary artery collateral circulation. *Circulation* 100:547–52.
- Schulz R, Mahmoudi S, Hattar K, Sibelius U, Olschewski H, Mayer K, Seeger W, Grimminger F. 2000a. Enhanced release of superoxide from polymorphonuclear neutrophils in obstructive sleep apnea. Impact of continuous positive airway pressure therapy. *Am J Respir Crit Care Med* 162:566–70.
- Schulz R, Schmidt D, Blum A, Lopes-Ribeiro X, Lucke C, Mayer K, Olschewski H, Seeger W, Grimminger F. 2000b. Decreased plasma levels of nitric oxide derivatives in obstructive sleep apnea: response to CPAP therapy. *Thorax* 55:1046–51.
- Shamsuzzaman AS, Gersh BJ, Somers VK. 2003. Obstructive sleep apnea: implications for cardiac and vascular disease. *JAMA* 290:1906–14.
- Smith PL, Gold AR, Meyers DA, Haponik EF, Bleecker ER. 1985. Weight loss in mildly to moderately obese patients with obstructive sleep apnea. *Ann Intern Med* 103(Pt 1):850–55.
- Smith R, Ronald J, Delaive K, Walld R, Manfreda J, Kryger MH. 2002. What are obstructive sleep apnea patients being treated for prior to this diagnosis? *Chest* 121:164–72.
- Song L, Leung C, Schindler C. 2001. Lymphocytes are important in early atherosclerosis. *J Clin Invest* 108:251–9.
- Stokes KY, Granger DN. 2005. Hypercholesterolemia: its impact on ischemia-reperfusion injury. *Expert Rev Cardiovasc Ther* 3:1061–70.
- Suzuki YJ, Jain V, Park AM, Day RM. 2006. Oxidative stress and oxidant signalign in obstructive sleep apnea and associated cardiovascular diseases. *Free Radic Biol Med* 40:1683–92.
- Tachibana N, Ayas NT, White DP. 2005. A quantitative assessment of sleep laboratory activity in the United States. *J Clin Sleep Med* 1:23–6.
- Ursavaş A, Karadağ M, Rodoplu E, Yilmaztepe A, Oral HB, Gözü RO. 2007. Circulating ICAM-1 and VCAM-1 levels in patients with obstructive sleep apnea syndrome. *Respiration* 74:525–32.
- Vanderlaan PA, Reardon CA. 2005. Thematic review series: the immune system and atherogenesis. The unusual suspects:an overview of the minor leukocyte populations in atherosclerosis. *J Lipid Res* 46:829–38.

- Vgontzas AN, Papanicolaou DA, Bixler EO, Hopper K, Lotsikas A, Lin HM, Kales A, Chrousos GP. 2000. Sleep apnea and daytime sleepiness and fatigue: relation to visceral obesity, insulin resistance, and hypercytokinemia. *J Clin Endocrinol Metab* 85:1151–8.
- Vinten-Johansen J. 2004. Involvement of neutrophils in the pathogenesis of lethal myocardial reperfusion injury. *Cardiovasc Res* 61:481–97.
- von der Thüsen JH, Kuiper J, van Berkel TJ, Biessen EA. 2003. Interleukins in atherosclerosis: molecular pathways and therapeutic potential. *Pharmacol Rev* 55:133–66.
- Wisse BE, Kim F, Schwartz MW. 2007. Physiology. An integrative view of obesity. *Science* 318:928–9.
- Yaggi HK, Concato J, Kernan WN, Lichtman JH, Brass LM, Mohsenin V. 2005. Obstructive sleep apnea as a risk factor for stroke and death. *N Engl J Med* 353:2034–41.
- Yamauchi M, Tamaki S, Tomoda K, Yoshikawa M, Fukuoka A, Makinodan K, Koyama N, Suzuki T, Kimura H. 2006. Evidence for activation of nuclear factor kappaB in obstructive sleep apnea. *Sleep Breath* 10:189–93.
- Young T, Palta M, Dempsey J, Skatrud J, Weber S, Badr S. 1993. The occurrence of sleep-disordered breathing among middle-aged adults. *N Engl J Med* 328:1230–5.
- Young T, Peppard P, Palta M, Hla KM, Finn L, Morgan B, Skatrud J. 1997. Population-based study of sleep-disordered breathing as a risk factor for hypertension. *Arch Intern Med* 157:1746–52.
- Young T, Peppard PE, Gottlieb DJ. 2002a. Epidemiology of obstructive sleep apnea: a population health perspective. *Am J Respir Crit Care Med* 165:1217–39.
- Young T, Shahar E, Nieto FJ, Redline S, Newman AB, Gottlieb DJ, Walsleben JA, Finn L, Enright P, Samet JM. Sleep Heart Health Study Research Group. 2002b. Predictors of sleep-disordered breathing in community-dwelling adults: the Sleep Heart Health Study. *Arch Intern Med* 162:893–900.
- Zamarrón-Sanz C, Ricoy-Galbaldon J, Gude-Sampedro F, Riveiro-Riveiro A. 2006. Plasma levels of vascular endothelial markers in obstructive sleep apnea. *Arch Med Res* 37:552–5.
- Zarbock A, Polanowska-Grabowska RK, Ley K. 2007. Platelet-neutrophil-interactions: linking hemostasis and inflammation. *Blood Rev* 21:99–111.
- Zidar N, Jeruc J, Balazic J, Stajer D. 2005. Neutrophils in human myocardial infarction with rupture of the free wall. *Cardiovasc Pathol* 14:247–50.